

Gangliosides Stabilize Ionic Homeostasis in Rat Cortical Synaptosomes Exposed to Toxic Concentrations of Glutamate

I. O. Zakharova, K. I. Shestak, V. G. Leont'ev, and N. F. Avrova

Translated from *Byulleten' Eksperimental'noi Biologii i Meditsiny*, Vol. 129, No. 1, pp. 45-47, January, 2000
Original article submitted December 8, 1998

Glutamate significantly increased intracellular calcium concentration, enhanced $^{45}\text{Ca}^{2+}$ entry, and activated $\text{Na}^+/\text{Ca}^{2+}$ exchanger in synaptosomes from rat cerebral cortex. Preincubation with GM_1 ganglioside reduced the glutamate-induced increase in intracellular calcium and $^{45}\text{Ca}^{2+}$ entry. Gangliosides and glutamate stimulated $\text{Na}^+/\text{Ca}^{2+}$ exchanger showing additive effects.

Key Words: *GM₁ ganglioside; glutamate; calcium; cerebral cortex synaptosomes*

Neuronal damage in ischemia, brain trauma, and some neurological diseases is largely determined by toxic effects of high concentrations of glutamate released into the intercellular space [2,3,12]. Excitotoxin-induced disturbance in intracellular calcium ($[\text{Ca}^{2+}]_i$) homeostasis is considered to be the major cause of neuronal death under these conditions [4].

Activation of N-methyl-D-aspartate receptors triggering this process is a necessary but not sufficient condition for sustained elevation of $[\text{Ca}^{2+}]_i$. An important role in the development of pathological processes initiated by excitotoxins is played by activation of free radical reactions [1,13] and other processes stimulating Ca^{2+} entry and inactivating the system stabilizing calcium homeostasis.

Gangliosides, the most complex animal glycosphingolipids, improve viability of cultured nervous cells exposed to glutamate [3,6] and significantly decrease $[\text{Ca}^{2+}]_i$, which seems to underlie their neuroprotective activity [12]. However, the mechanism of this effect remains unclear. We found no reports on the effects of gangliosides on glutamate-induced activation of Ca^{2+} entry or activity of $\text{Na}^+/\text{Ca}^{2+}$ exchanger.

Our aim was to study the mechanism of stabilizing effect of gangliosides on Ca^{2+} homeostasis dis-

turbed by excessive glutamate by investigating the effects of glutamate and gangliosides on $[\text{Ca}^{2+}]_i$, $^{45}\text{Ca}^{2+}$ entry, and activity of $\text{Na}^+/\text{Ca}^{2+}$ exchanger in synaptosomes from rat cerebral cortex.

MATERIALS AND METHODS

Synaptosomes isolated from the brain of male Wistar rats (150-180 g) as described elsewhere [9] were resuspended in buffer A containing (in mM): 135 NaCl, 1.2 MgSO_4 , 2.0 KCl, 10 glucose, and 20 tris-HCl (pH=7.4), centrifuged at 15,000g for 15 min, and resuspended again in buffer A (4-5 mg/ml protein concentration).

$[\text{Ca}^{2+}]_i$ was measured with Fura-2M (Sigma), a fluorescent calcium probe. Synaptosomes (1 ml) were mixed with 1 ml buffer A containing 0.5% BSA (free from fatty acids) and 8 μM Fura-2M dissolved in 0.4% DMSO and incubated for 40 min at 30°C [16]. Preincubation with 50 nM GM_1 ganglioside was performed simultaneously with Fura-2M loading, which was stopped by adding 20-fold volume of buffer A followed by 15-min centrifugation at 12,000g. The sediment was resuspended in MgSO_4 -free buffer B containing (in mM): 135 NaCl, 2.0 KCl, 1.0 CaCl_2 , 10 glucose, and 20 tris-HCl (pH=7.4) and incubated with glutamate and other test compounds for 15 min at 30°C. Fluorescence was measured on a MPF-2A fluorimeter (Hitachi) at 340 and 380 nm excitation and 510 nm

Laboratory of Comparative Neurochemistry, I. M. Sechenov Institute of Evolutionary Physiology and Biochemistry, St. Petersburg. **Address for correspondence:** avrova@lipid.mail.iephb.ru. Avrova N. F.

emission wavelength. $[Ca^{2+}]_i$ was calculated according to the formula:

$$[Ca^{2+}]_i = K_d \times (R - R_{\min} / R_{\max} - R) \times Sf_2 / Sb_2, \text{ where}$$

R is the ratio of Fura-2 fluorescences at 340 and 380 nm; R_{\min} and R_{\max} are the same ratios in Ca^{2+} -free medium and after saturation with Ca^{2+} , respectively. Sf_2 is fluorescence at 380 nm in Ca^{2+} -free medium and Sb_2 is the same after saturation with Ca^{2+} . R_{\min} and Sf_2 were measured in the presence of 2 mM EGTA and 0.1% Triton X-100; R_{\max} and Sb_2 were measured after addition of 5 mM Ca^{2+} to saturate both EGTA and Fura-2. K_d was taken as 184 nM at 30°C [8].

Calcium accumulation in the control and glutamate-exposed synaptosomes was measured after 30-sec incubation in a medium containing (in mM): 135 NaCl, 1.2 K_2HPO_4 , 3.0 KCl, 1.0 $CaCl_2$, 10 glucose, 20 tris-HCl (pH=7.4), and $^{45}Ca^{2+}$ (54 μ Ci/mmol). The protein content was about 200 μ g/ml. Preincubation with various concentrations of GM_1 ganglioside was performed for 1 h at 37°C. Activity of Na^+/Ca^{2+} exchanger was determined by the difference in $^{45}Ca^{2+}$ accumulation in NaCl- and choline chloride-containing media [5]. Radioactivity was measured on an LKB scintillation counter.

GM_1 ganglioside was isolated on silica gel columns [15]. Gangliosides were isolated from bovine brain as described elsewhere [7]. The isolated GM_1 was of 98-99% purity as shown by high-performance thin layer chromatography on Merck plates.

Synaptosomal protein concentration was measured [10] using bovine serum albumin as the standard. The data were analyzed statistically using Student's t test.

RESULTS

Glutamate (1 mM) significantly elevated synaptosomal $[Ca^{2+}]_i$ by on average $43.6 \pm 8.6\%$ ($p < 0.05$, Table 1). This finding agrees with published data showing that exposure to glutamate increases $[Ca^{2+}]_i$ in cultured cortical, hippocampal, and other neurons as well as in rat hippocampal synaptosomes [11,13,14]. Preincubation with 50 nM GM_1 ganglioside prevented the increase in $[Ca^{2+}]_i$ induced by glutamate alone or in combination with gangliosides ($p < 0.05$, Table 1).

Glutamate significantly increased $^{45}Ca^{2+}$ entry into rat brain synaptosomes ($p < 0.01$). Preincubation with GM_1 ganglioside inhibited this effect (Table 2). This inhibition was dose-dependent within the concentration range from 1 pM to 10 nM. The most pronounced inhibition of calcium response to glutamate was attained with 10 nM GM_1 ($p < 0.01$), when $^{45}Ca^{2+}$ entry in the presence of glutamate did not differ from the control. GM_1 also exerted a significant, but less pro-

TABLE 1. Effect of Glutamate and GM_1 Ganglioside on $[Ca^{2+}]_i$ in Cortical Synaptosomes from Rat Brain ($M \pm m$)

Experiment	$[Ca^{2+}]_i$, nM
Control ($n=14$)	501.6 ± 64.1
Glutamate, 1 mM ($n=14$)	$720.2 \pm 103.7^*$
GM_1 , 50 nM ($n=5$)	574.4 ± 103.9
Glutamate, 1 mM and GM_1 , 10 nM ($n=5$)	$1074.6 \pm 117.3^*$
GM_1 , 50 nM+glutamate, 1 mM, and GM_1 , 10 nM ($n=5$)	$635.0 \pm 133.9^{**}$

Note. * $p < 0.05$ in comparison with the control; ** $p < 0.05$ in comparison with glutamate alone.

TABLE 2. Effect of Glutamate and GM_1 Ganglioside on $^{45}Ca^{2+}$ Entry into Cortical Synaptosomes from Rat Brain ($M \pm m$)

Experiment	$^{45}Ca^{2+}$ entry, nmol/mg protein/30 sec
Control	2.63 ± 0.27
Glutamate, 1 mM	$7.87 \pm 1.54^*$
GM_1 , 1 pM+glutamate	7.19 ± 2.06
GM_1 , 10 pM+glutamate	6.91 ± 1.93
GM_1 , 1 nM+glutamate	5.18 ± 0.61
GM_1 , 10 nM+glutamate	$2.32 \pm 0.16^*$
GM_1 , 100 nM+glutamate	$4.54 \pm 0.52^{**}$
GM_1 , 1 μ M+glutamate	4.24 ± 0.19
GM_1 , 10 μ M+glutamate	$4.52 \pm 0.39^{**}$

Note. * $p < 0.01$ in comparison with the control; * $p < 0.01$, ** $p < 0.05$ in comparison with the effect of glutamate alone (Student's t test).

TABLE 3. Effect of Glutamate and GM_1 Ganglioside on the Activity of Na^+/Ca^{2+} Exchanger in Rat Brain Synaptosomes

Experiment	Activity of Na^+/Ca^{2+} exchanger, %
Control	100
Glutamate, 1 mM	192.9 ± 42.8
GM_1 , 40 pM	183.5 ± 41.3
GM_1 , 40 pM+glutamate	$224.9 \pm 39.4^*$

Note. * $p < 0.05$ in comparison with the control.

nounced inhibitory effect in a concentration of 10 μ M corresponding to a micellar form of the ganglioside. The effects of glutamate and gangliosides on Na^+/Ca^{2+} exchanger were similar and additive: significant activation of Na^+/Ca^{2+} exchange (0.64 ± 0.03 nmol Ca^{2+} /mg protein/30 sec) was observed only after combined exposure to glutamate and GM_1 (Table 3).

These data suggest that the ability of ganglioside to inhibit Ca^{2+} entry induced by glutamate and other excitotoxins plays an important role in the stabiliza-

tion of intracellular Ca^{2+} homeostasis. As previously shown [6,12] gangliosides do not interfere with glutamate-induced activation of N-methyl-D-aspartate receptors and exert their effect at later stages. The increase in $[\text{Ca}^{2+}]_i$ after exposure to toxic glutamate concentrations can result from increased membrane permeability due to accumulation of lipid peroxidation products [14]. Glutamate-induced stimulation of $^{45}\text{Ca}^{2+}$ entry is significantly inhibited by α -tocopherol and SOD [1], therefore the effect of gangliosides can be associated with inhibition of free radical reactions activated by glutamate [2,15]. Activation of $\text{Na}^+/\text{Ca}^{2+}$ exchanger in rat brain synaptosomes by glutamate and GM_1 can also have a protective effect, since its inactivation significantly promotes neuronal death in culture [3].

The study was supported by the Russian Foundation for Basic Research (grant No. 98-04-49686).

REFERENCES

1. V. A. Tyurin, T. V. Sokolova, V. O. Gonchar, *et al.*, *Zh. Evoluts. Biokhim.*, **34**, No. 5, 325-332 (1998).
2. N. F. Avrova, I. V. Victorov, V. A. Tyurin, *et al.*, *Neurochem. Res.*, **23**, 947-954 (1998).
3. N. Andreeva, B. Khodorov, E. Stelmaschook, *et al.*, *Brain Res.*, **248**, 322-325 (1991).
4. D. W. Choi and S. M. Rotman, *Ann. Rev. Neurosci.*, **13**, 171-182 (1990).
5. O. P. Couninho, C. A. Carvalho, and A. P. Carvalho, *Brain Res.*, **290**, 261-271 (1984).
6. M. Favaron, H. Manev, H. Alho, *et al.*, *Proc. Natl. Acad. Sci. USA*, **85**, 7351-7355 (1988).
7. J. Folch, M. Lees, and G. S. Sloan-Stanley, *J. Biol. Chem.*, **226**, 497-509 (1957).
8. G. Grynkiewicz, M. Poenie, and R. Tsien, *Ibid.*, **260**, 3440-3450 (1985).
9. F. Hajos, *Brain Res.*, **93**, 485-489 (1975).
10. O. H. Lowry, N. J. Rosenbrough, A. L. Farr, and R. J. Randall, *J. Biol. Chem.*, **193**, 265-275 (1951).
11. J. O. Malva, A. F. Ambrosio, and A. P. Carvalho, *Neurochem. Int.*, **32**, 7-16 (1998).
12. H. Manev, M. Favaron, A. Guidotti, and E. Costa, *Mol. Pharmacol.*, **36**, 106-112 (1989).
13. M. P. Mattson, M. A. Lovell, K. Furukawa, and W. R. Markesbery, *J. Neurochem.*, **65**, 1740-1751 (1995).
14. L. Tretter and V. Adam-Vizi, *Ibid.*, **66**, 2057-2066 (1995).
15. V. A. Tyurin, Y.Y. Tyurina, and N. F. Avrova, *Neurochem. Int.*, **20**, 401-407 (1992).
16. S. L. Yates, E. N. Fluhler, and P. M. Lippicello, *J. Neurosci. Res.*, **32**, 255-260 (1992).